

Validation of Commercial tool Antibodies

The Antibody Society Webcast series – Antibody Validation #5

Fit-for-purpose Positive and Negative controls

Simon L. Goodman

Science and Technology Program Manager

The Antibody Society

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Antibody Validation: a 9-part series

1. Andreas Pluckthun : The different antibody formats
2. Glenn Begley : Antibodies and the reproducibility crisis in biological science
Cecilia Williams : The ErbB story – is your antibody like this?
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- 5. Giovanna Roncador: : Correct positive and negative controls in validation**
6. Aldrin Gomes : Standard technology: “even” Western blots are non-trivial
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Antibody Validation

The Antibody Society Webcast series - #5 **Fit-for-Purpose Positive and Negative controls**

Giovanna Roncador

Head of the Monoclonal Antibodies Unit

Chair of EuroMabNet

CNIO, Spain

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Giovanna Roncador

CNIO Madrid, EuroMabNet

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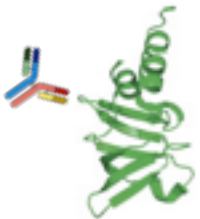
What is the goal of antibody validation?



Is to demonstrate that the Ab is **specific**, **selective**, and **reproducible** in the application in which it is going to be used.

Specificity

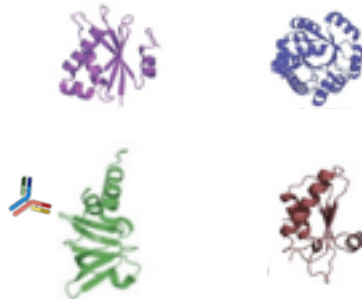
capability to bind specifically to one **unique epitope**



+

Selectivity

ability of an Ab to react only with **one antigen**



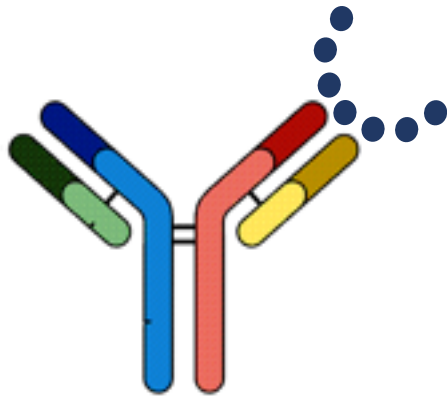
+

Reproducibility

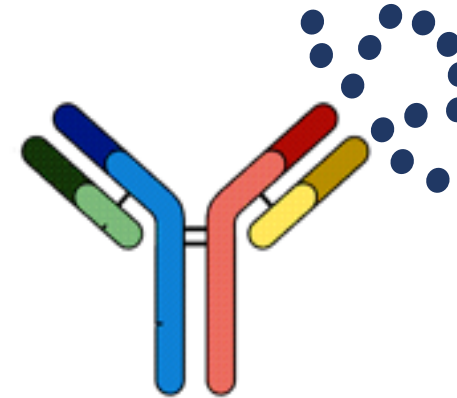
ability to **duplicate results** over long periods of time by different laboratories.



Ab performances is application-specific

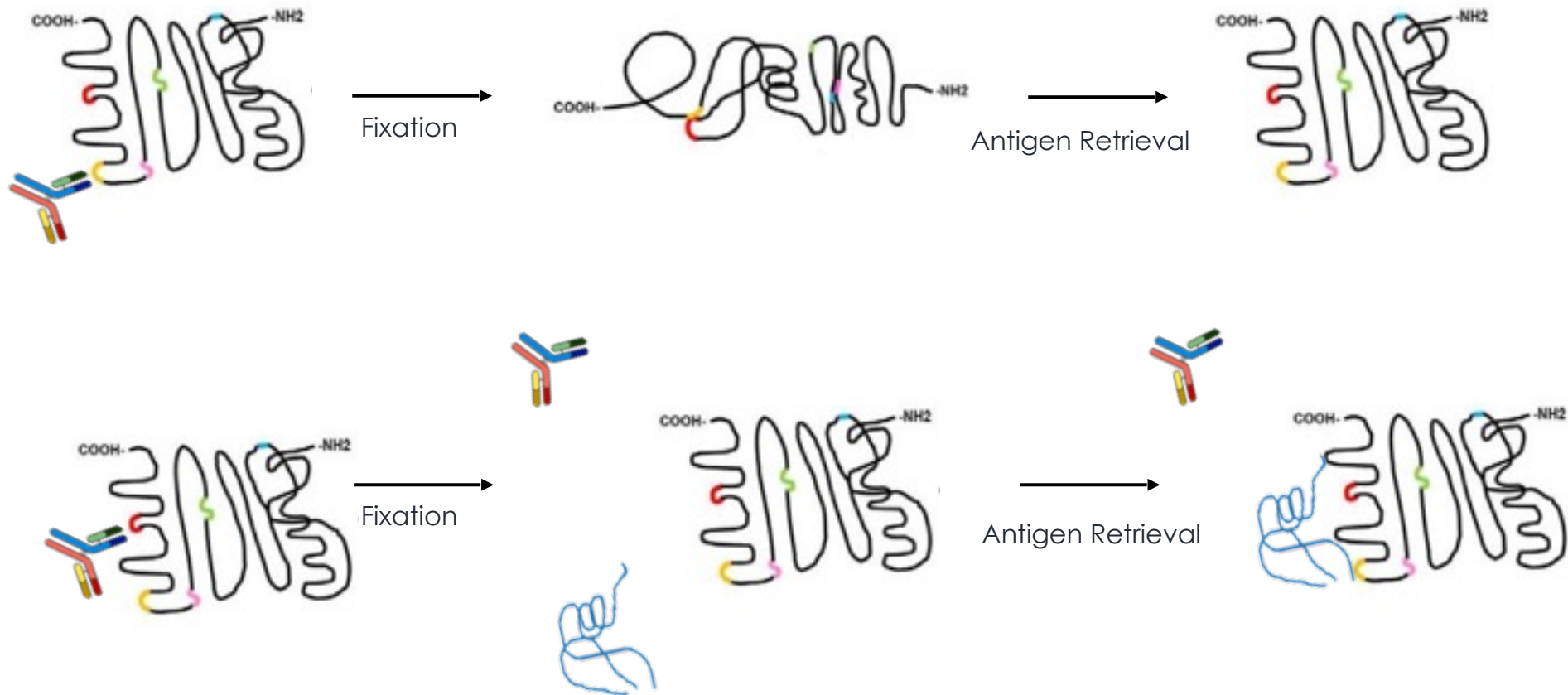


Linear Epitopes Western
Blotting, IF, IHC



Conformational Epitopes
ELISA , Flow Cytometry, IP

Conformational and linear epitopes of native/partially-unfolded proteins (fixed & cross-linked)



Before starting Ab Validation: review your target



Full literature review
of your target

- ➔ Build a picture of **when** and **where** expression is expected
(expected sub cellular localization)
- ➔ Discover possible cross reactivity
with related proteins
- ➔ Identify all known variants
 - alternative splicing,
 - proteolytic cleavage
 - post-translational modification

Importance of +/- controls

The **key** to proving antibody specificity is often the correct use of controls

Positive control:

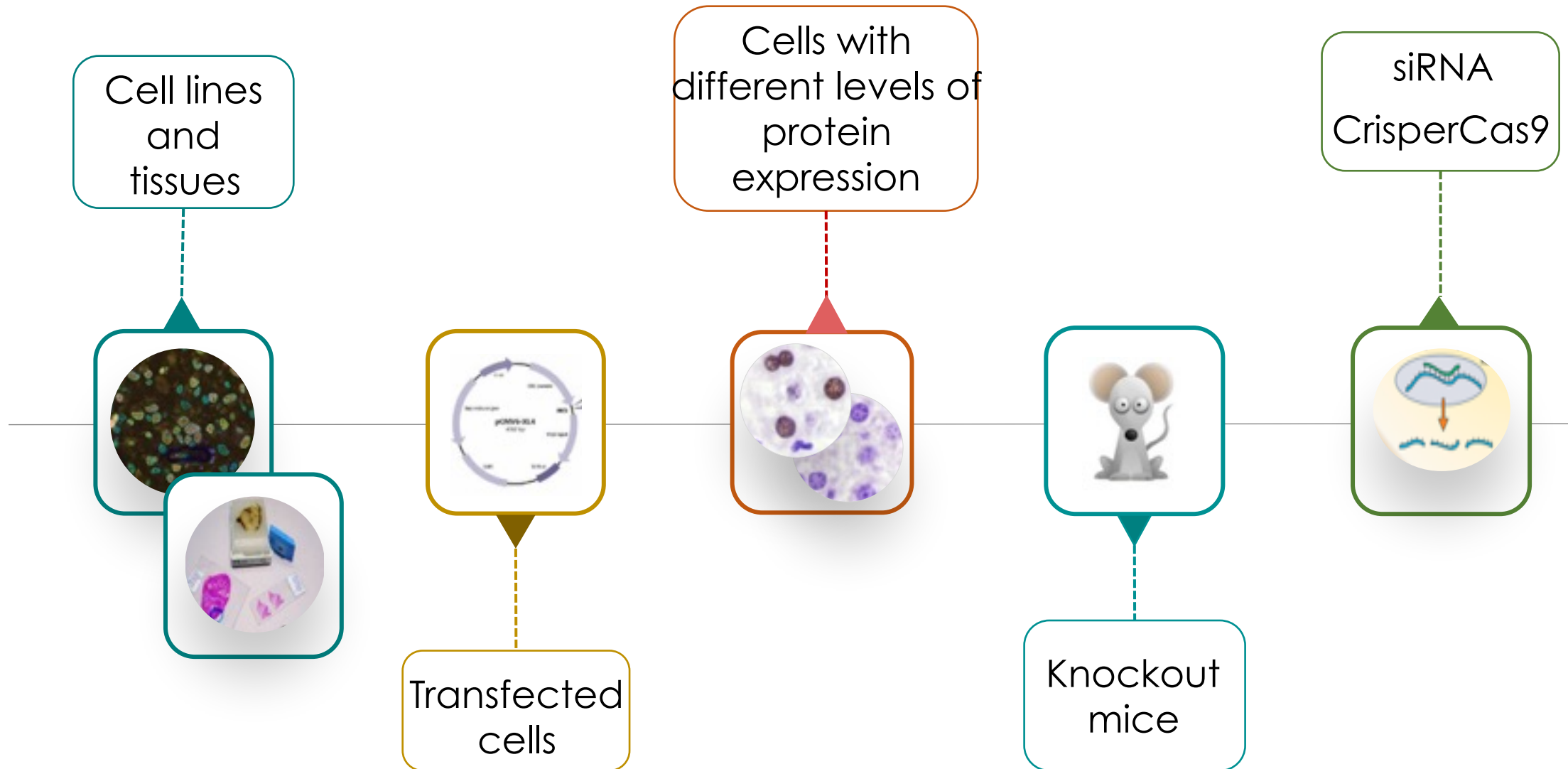
should confirm that your target antigen is expressed on cells and tissues



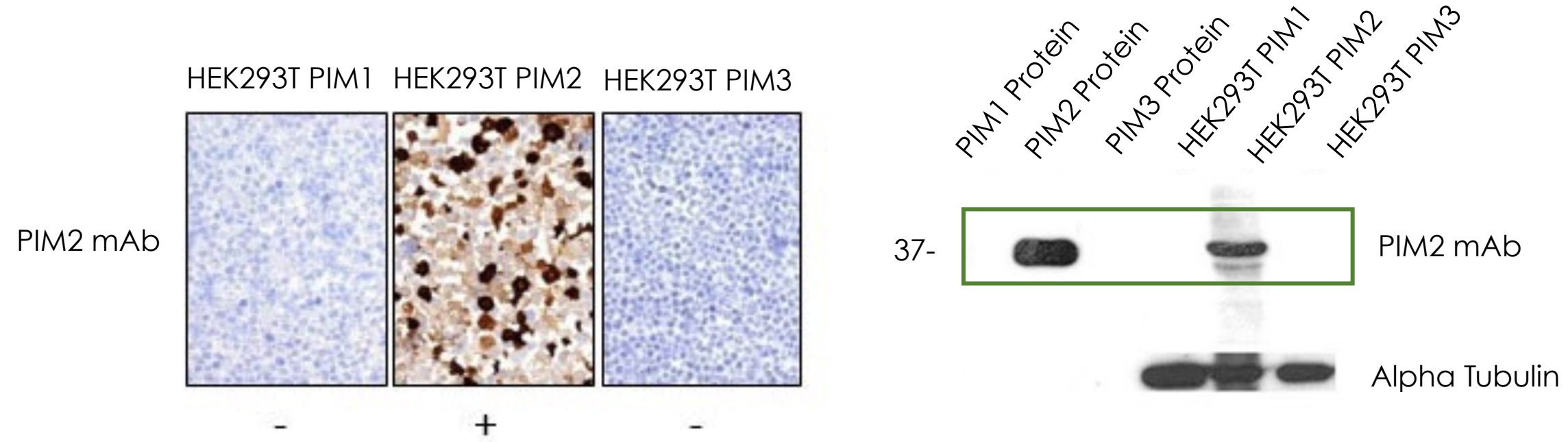
Negative control:

should consist of tissues or cells where your target protein is known to be absent

Positive and Negative controls

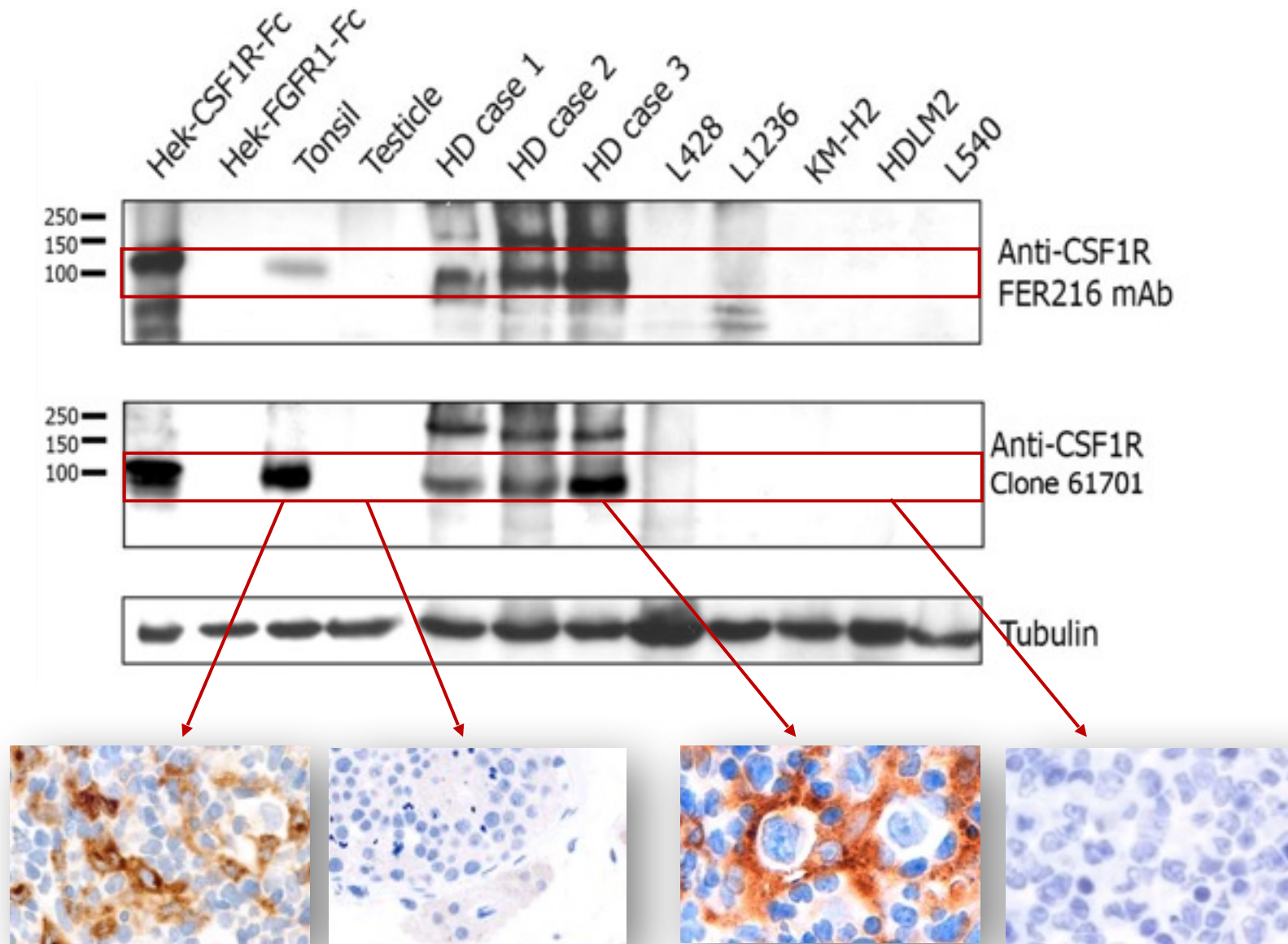


Transfected cells are a useful tool to check Ab cross reactivity

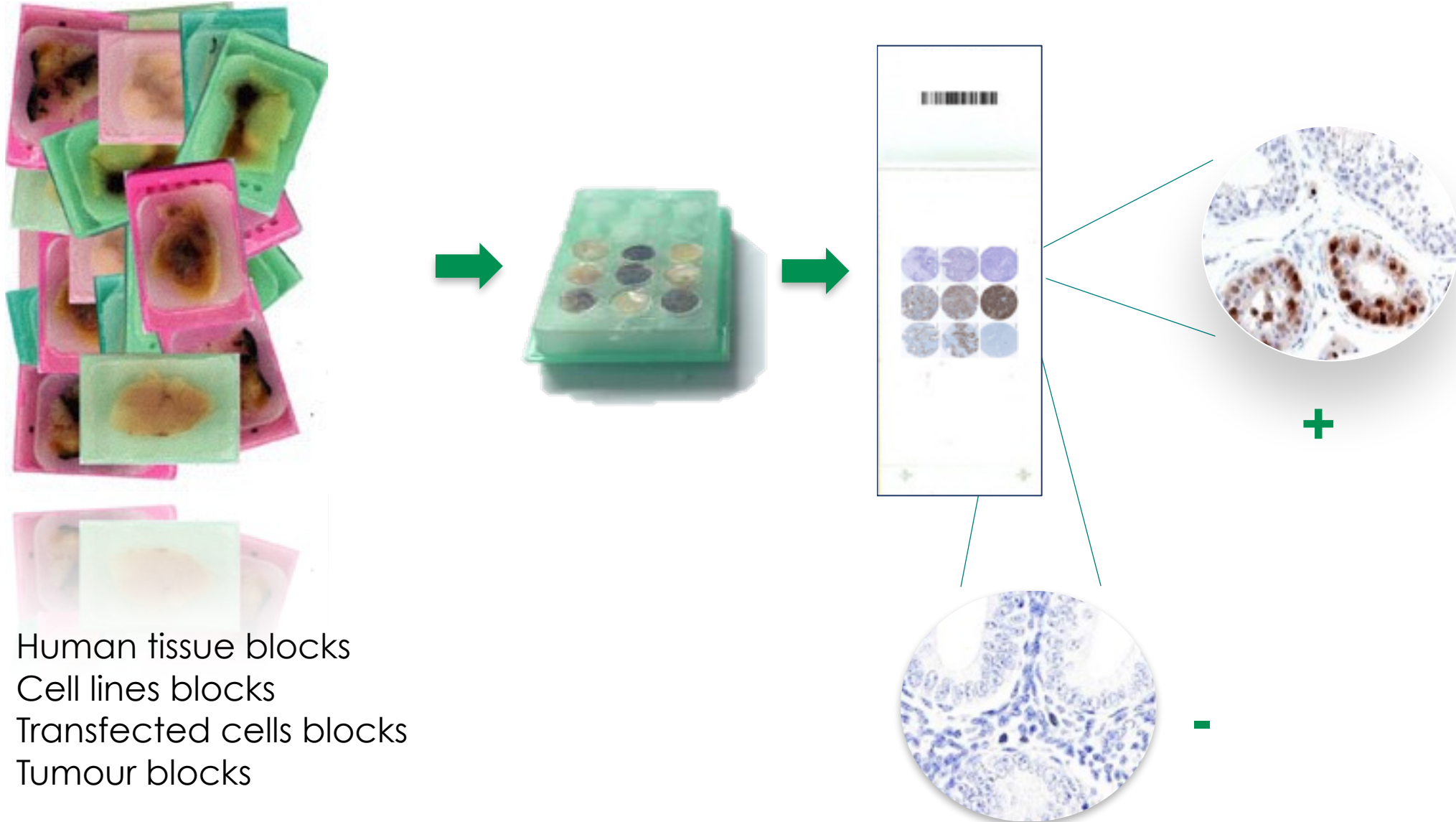


In some cases Abs that recognise it target in transfected cells may still be unable to detect the native endogenously expressed protein

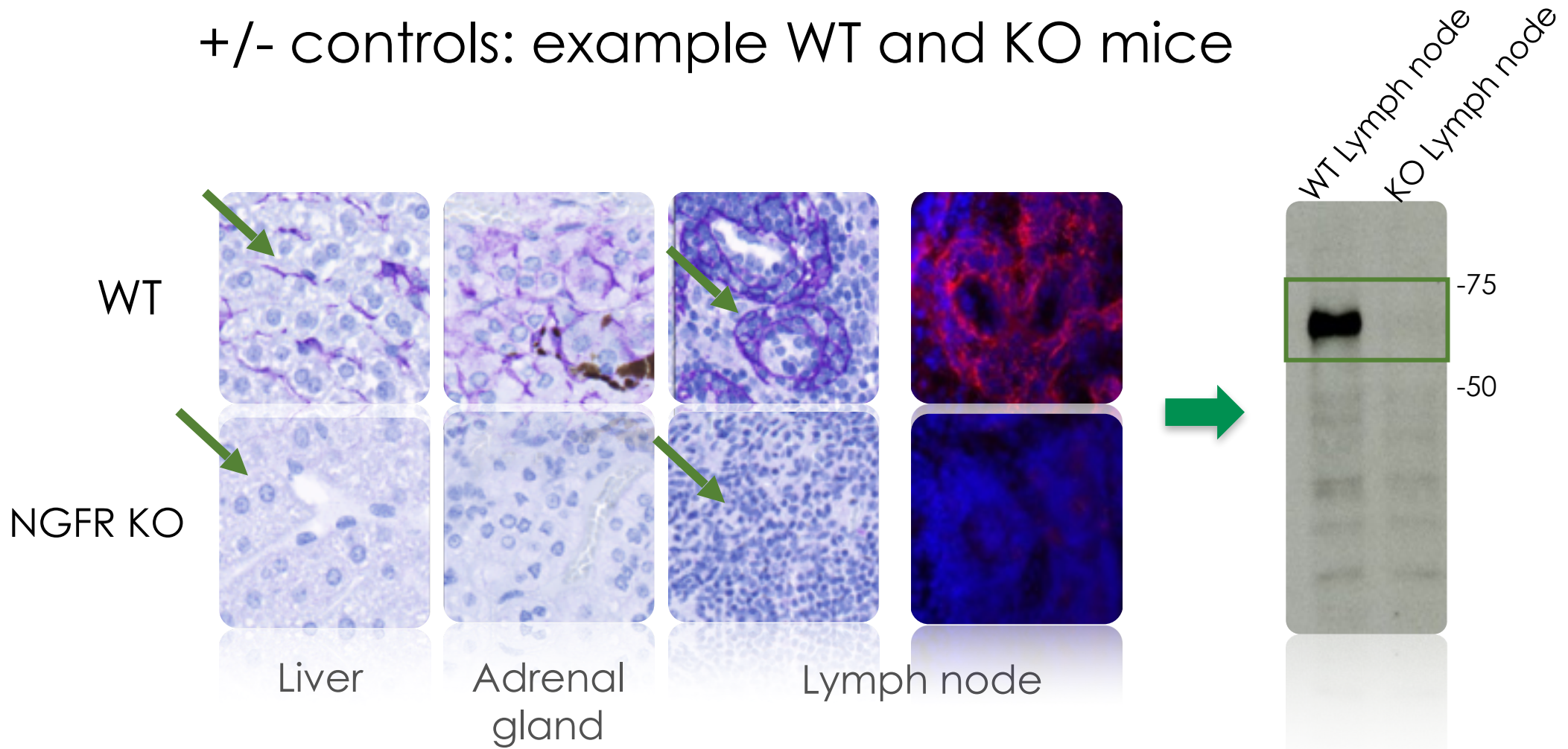
+/- controls: example Western Blotting



Small tissue arrays are a useful tool for Ab validation by IHC



+/- controls: example WT and KO mice

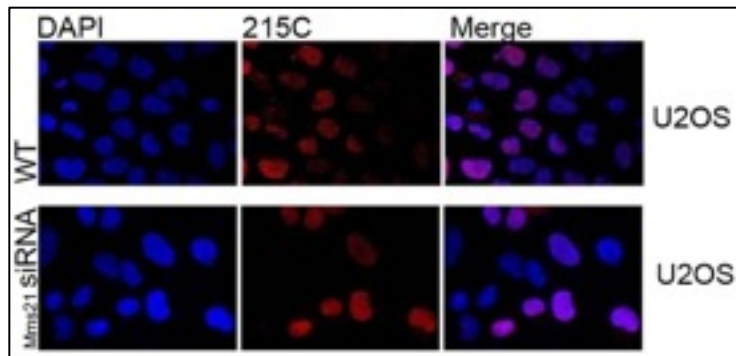
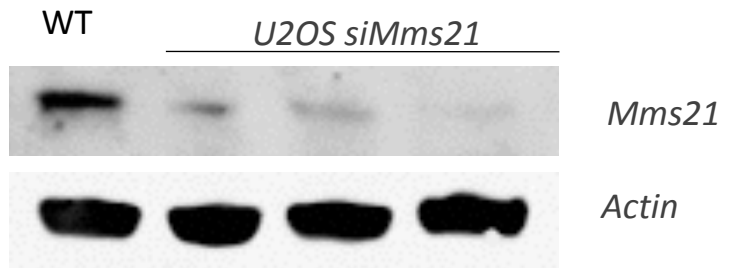


The use of KO mice is only applicable if your antibody recognizes a mouse molecule.

+/- controls: knock down and knock out methods

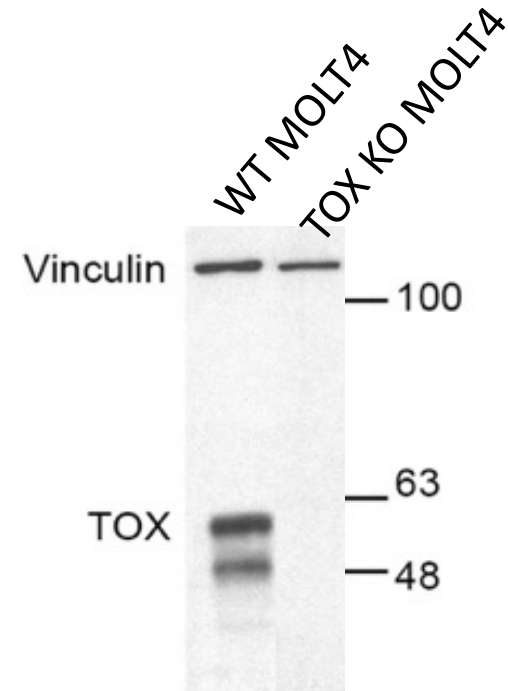
Example siRNA:

Ab against Mms21

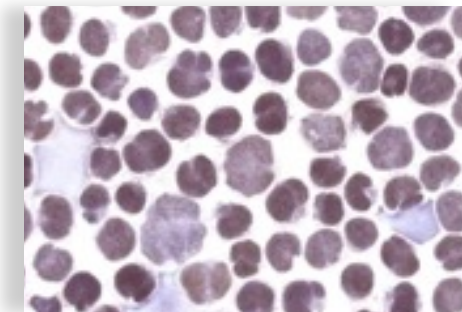


Example CrisperCas9:

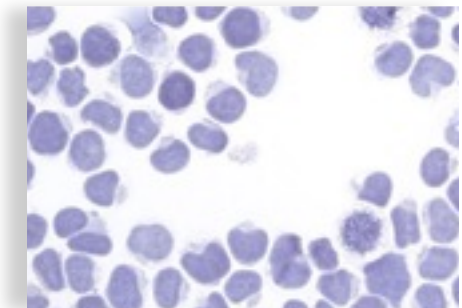
Ab against TOX



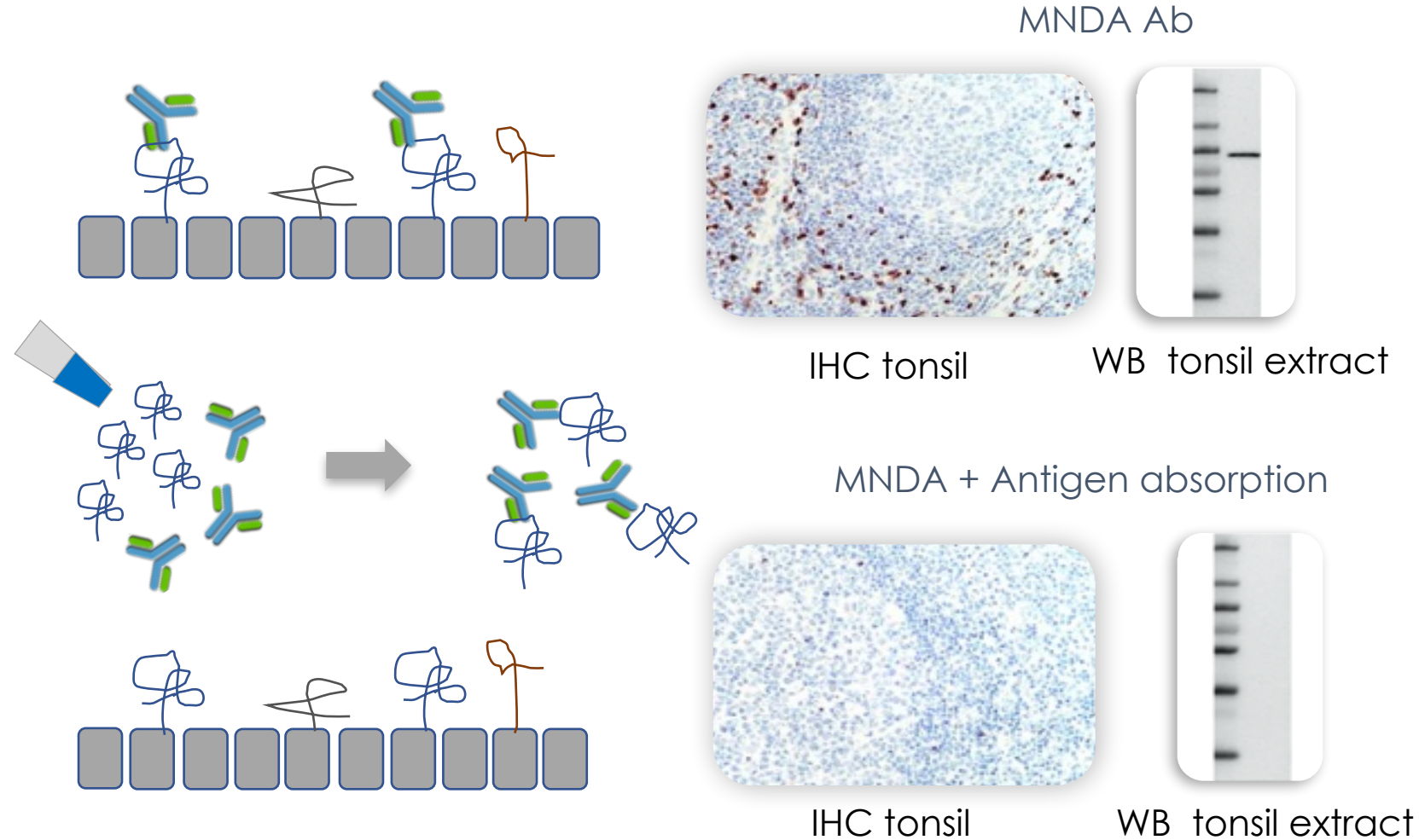
WT MOLT4



TOX KO MOLT4



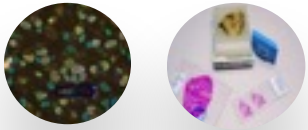
Antigen Preadsorption do not determine specificity



Blocking reactivity with excess antigen demonstrates that the reactivity is produced by an antibody that recognizes that antigen and **DO NOT** prove specificity.

Summary

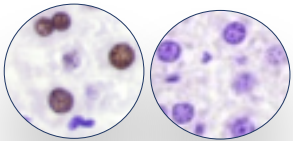
Cell lines and tissues



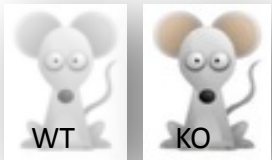
Transfected cells



different levels
of protein expression



Knockout (KO) mice



Haploid KO cell models



Use more than one
Abs
against your target

IHC

WB

IP

Flow
Cytometry

CHIP

Other
Techniques

Use more than one technique

Confirm that the
result obtained fit
with how described
in the literature

The validation
data should be
available to the
scientific
community.

Publications

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Next Webcast in Antibody Validation: a 9-part series

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Presented by Giovanna Roncador

Produced and Directed by Simon L. Goodman

Production Manager Fran Breden

Written by Simon Goodman

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Administrative Support: Dr. Fran Breden and Dr. Mini Muralidharan

Executive Director: Dr. Jan Reichert

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